

# Sensitivity of the Human Circadian System to Short-Wavelength (420-nm) Light

George C. Brainard,<sup>\*1</sup> David Sliney,<sup>†,2</sup> John P. Hanifin,<sup>\*</sup> Gena Glickman,<sup>\*,3</sup> Brenda Byrne,<sup>\*</sup> Jeffrey M. Greeson,<sup>\*,4</sup> Samar Jasser,<sup>\*,5</sup> Edward Gerner,<sup>\*</sup> and Mark D. Rollag<sup>\*</sup>

<sup>\*</sup>*Department of Neurology, Thomas Jefferson University, Philadelphia, PA 19107, USA,*

<sup>†</sup>*Laser/Optical Radiation Program, US Army Center for Health Promotion and Preventive Medicine, Aberdeen Proving Ground, MD 21010, USA*

*Abstract* The circadian and neurobehavioral effects of light are primarily mediated by a retinal ganglion cell photoreceptor in the mammalian eye containing the photopigment melanopsin. Nine action spectrum studies using rodents, monkeys, and humans for these responses indicate peak sensitivities in the blue region of the visible spectrum ranging from 459 to 484 nm, with some disagreement in short-wavelength sensitivity of the spectrum. The aim of this work was to quantify the sensitivity of human volunteers to monochromatic 420-nm light for plasma melatonin suppression. Adult female ( $n = 14$ ) and male ( $n = 12$ ) subjects participated in 2 studies, each employing a within-subjects design. In a fluence-response study, subjects ( $n = 8$ ) were tested with 8 light irradiances at 420 nm ranging over a 4-log unit photon density range of  $10^{10}$  to  $10^{14}$  photons/cm<sup>2</sup>/sec and 1 dark exposure control night. In the other study, subjects ( $n = 18$ ) completed an experiment comparing melatonin suppression with equal photon doses ( $1.21 \times 10^{13}$  photons/cm<sup>2</sup>/sec) of 420 nm and 460 nm monochromatic light and a dark exposure control night. The first study demonstrated a clear fluence-response relationship between 420-nm light and melatonin suppression ( $p < 0.001$ ) with a half-saturation constant of  $2.74 \times 10^{11}$  photons/cm<sup>2</sup>/sec. The second study showed that 460-nm light is significantly stronger than 420-nm light for suppressing melatonin ( $p < 0.04$ ). Together, the results clarify the visible short-wavelength sensitivity of the human melatonin suppression action spectrum. This basic physiological finding may be useful for optimizing lighting for therapeutic and other applications.

*Key words* melatonin, action spectrum, circadian, wavelength, light, pineal gland, neuroendocrine, photoreception

1. To whom all correspondence should be addressed: George C. Brainard, Department of Neurology, Thomas Jefferson University, 1025 Walnut Street, Suite 507, Philadelphia, PA 19107; e-mail: george.brainard@jefferson.edu.
2. Present address: Consulting Medical Physicist, 406 Streamside Drive, Fallston, MD 21047-2806.
3. Present address: Department of Psychology, University of California, San Diego, 9500 Gilman Drive, La Jolla, CA 92093-0109.
4. Present address: Duke Integrative Medicine, Duke University Medical Center, DUMC Box 102904, Durham, NC 27710.
5. Present address: University of Pennsylvania, Department of Psychiatry, 3535 Market Street, 2nd Floor, Philadelphia, PA 19104.

JOURNAL OF BIOLOGICAL RHYTHMS, Vol. XX No. X, Month XXXX xx-xx

DOI: 10.1177/0748730408323089

© 2008 Sage Publications

Light can be a potent therapeutic intervention for patients with selected affective and sleep disorders as well as healthy individuals who have circadian disruption due to shift work, transcontinental jet travel, or manned space flight (Commission Internationale de l'Éclairage [CIE], 2004, 2006; Dijk et al., 2001). The ocular photoreceptive physiology that supports the therapeutic capacity of light, however, has been described only in a nascent fashion.

Two action spectra identified 446 to 477 nm as the most potent wavelength region for acute plasma melatonin suppression in human subjects (Brainard et al., 2001; Thapan et al., 2001). Data from both studies shared many similarities and suggested that a novel vitamin A retinaldehyde-based photopigment was primarily responsible for this effect. There was poor agreement between these studies, however, on the sensitivity to light at 420 to 424 nm. One action spectrum tested subjects with 420 nm at 2 different intensities and calculated that 420 nm would be substantially weaker than the peak wavelength in the melatonin action spectrum (Brainard et al., 2001). In contrast, the other action spectrum tested 424 nm at 4 intensities and showed that 424 nm was substantially stronger than the peak wavelength of its action spectrum (Thapan et al., 2001).

Recently, there has been an upheaval in understanding the photoreceptive input to the circadian system. A new photopigment, named *melanopsin*, has been localized in the retinas of rodents, monkeys, and humans (Provencio et al., 2000). Melanopsin is found in a specific subtype of the retinal output neuron, the intrinsically photosensitive ganglion cells (ipRGCs) that project to the SCN (Berson et al., 2002; Hattar et al., 2002; Gooley et al., 2001). The ipRGC responses to light appear to parallel those of melatonin suppression and photic entrainment, suggesting that these cells are primary photoreceptors involved in circadian regulation (Berson et al., 2002; Hattar et al., 2002).

Altogether, 9 analytic action spectra in humans, monkeys, and rodents have demonstrated the wavelength sensitivity of physiological responses that are mediated by the newly characterized ipRGCs (for review, see Brainard et al., 2001; Thapan et al., 2001; Hattar et al., 2002; Hattar et al., 2003; Dacey et al., 2005; Gamlin et al., 2007; Brainard and Hanifin, 2005). Notably, all of the action spectra were fit to single-opsin nomograms with high coefficients of correlation and identify shorter wavelength peak photosensitivities in the blue region of the visible

spectrum with calculated peaks ranging from 459 to 484 nm. Despite differences in laboratories, physiological endpoints, animal models, and specific techniques, there is a consistent detection of peak responses in the blue spectrum (for review, see Brainard and Hanifin, 2005). Together, these studies suggest that a novel ocular photoreceptor system is involved in phototransduction for circadian, neuroendocrine, and other neurobehavioral responses (such as pupil constriction, acute alerting effects, cognitive responses, etc.).

Three recent studies have provided compelling evidence that melanopsin is the photopigment that mediates ipRGC phototransduction (Melyan et al., 2005; Qiu et al., 2005; Panda et al., 2005). Specifically, when mouse cells are transfected with the human melanopsin gene, they become photosensitive with peak response deeper into the violet-indigo portion of the visible spectrum in the range of 360 to 430 nm (Melyan et al., 2005). That pattern of photosensitivity encompasses the *in vitro* peak absorption spectrum of melanopsin (Newman et al., 2003). In contrast, human kidney cells transfected with the mouse melanopsin gene are light responsive with peak sensitivity at 479 nm (Qiu et al., 2005). Likewise, the expression of mouse melanopsin in *Xenopus* oocytes confers peak sensitivity at 480 nm (Panda et al., 2005). These latter 2 studies identify peaks that are reflective of the action spectra that peak in the blue spectrum (for review, see Gamlin et al., 2007; Brainard and Hanifin, 2005). Clearly, there is not complete agreement in specific wavelength sensitivity across the action spectra studies, the study on melanopsin absorption spectrum, and the photic responses of cells transfected with the melanopsin gene. The collective results are consistent, however, in demonstrating a wavelength signature in the short-wavelength visible spectrum that appears distinct from the wavelength sensitivity of rod and cone systems that mediate vision.

The aim of this study was to quantify the sensitivity to monochromatic 420 nm for pineal melatonin suppression in humans. The data demonstrate that there is a fluence-response relationship between 420-nm light and melatonin suppression that is consistent with the fluence-response curves for 8 other wavelengths (Brainard et al., 2001). A second study shows that 460-nm light is approximately twice as strong as 420-nm light for suppressing plasma melatonin. Together, the results clarify the short visible wavelength sensitivity of the human melatonin suppression action spectrum.

## RESEARCH DESIGN AND METHODS

### Study Designs

In the first study, 8 subjects completed a within-subjects fluence-response experiment that tested 8 light irradiances at 420 nm and 1 dark exposure control night on nocturnal melatonin suppression. In the second study, 18 subjects completed a within-subjects experiment comparing melatonin suppression with equal photon doses of 420-nm and 460-nm monochromatic light and a dark exposure control night.

### Subjects

The healthy subjects in both studies had a mean  $\pm$  SEM age of  $24.5 \pm 0.6$  years, demonstrated normal color vision by the Ishihara test, had a mean wakeup time of  $6:54 \text{ AM} \pm 18 \text{ min}$ , and signed an approved institutional review board (IRB) consent document. All subjects in the fluence-response study also demonstrated normal color vision by their Farnsworth Munsell D-100 color vision score (mean  $\pm$  SEM of  $47.1 \pm 6.8$ ). Five women and 3 men were in the dose-response study. Nine women and 9 men were in the 420/460-nm comparison study.

### Light Exposure Protocol

As described in detail elsewhere (Brainard et al., 2001), each experiment began at midnight when subjects had their pupils dilated with 0.5% cyclopentolate, were blindfolded, and remained awake and sitting upright in darkness for 120 min. While blindfolded, a blood sample was taken just prior to 2:00 am, and subjects were then exposed to a 90-min light stimulus until 3:30 am. During light exposure, each subject sat quietly with his or her eyes open and his or her head resting in an ophthalmologic head holder facing a patternless, white Ganzfeld apparatus encompassing the entire visual field. At 3:30 am, a second blood sample was taken. Each subject was exposed to complete darkness from 2:00 to 3:30 am on the control night. There were at least 6 days between each nighttime test. Melatonin was quantified with a radioimmunoassay (RIA) with a minimum detection limit of 0.5 to 5.0 pg/mL (Brainard et al., 2001). RIA control samples had 14% and 22% interassay coefficients of variation.

### Light Production and Measurement

As detailed elsewhere, monochromatic wavelengths were produced by arc lamps collimated into

a grating monochromator (Brainard et al., 2001). The resulting light beam was directed into the top area of a Ganzfeld dome and reflected evenly off the dome surface into volunteers' eyes. Half-peak bandwidths of the monochromatic stimuli were 14 nm for the dose-response study and 10 nm for the comparison study. Wavelengths at the level of subjects' corneas were measured with a portable spectroradiometer (Ocean Optics S2000). Routine measurement of the light irradiance ( $\mu\text{W}/\text{cm}^2$ ) was done with both a Tektronix J16 Radiometer/Photometer with a J6512 irradiance probe, which was not cosine corrected, and an International Light 1400A with an SEL033 #6857 detector head and an F #23102 filter and cosine correction. Each of these meters was calibrated annually and was benchmarked to a reference meter (EG & G Model 580-23A Detector) at the Laser/Optical Radiation Program (Aberdeen Proving Ground, MD). All spectroradiometric and radiometric equipment was calibrated with a standard lamp traceable to the National Institute of Standards and Technology (NIST). Experimental light stimuli were measured at volunteers' eye level immediately before and after the 90-min exposure. In the 420-nm fluence-response study, intensities covered a 4-log unit photon density range of  $10^{10}$  to  $10^{14}$  photons/ $\text{cm}^2$ . In the study comparing 420 and 460 nm, the photon density was  $1.21 \times 10^{13}$  photons/ $\text{cm}^2/\text{sec}$ . An earlier study, which measured mean transmittance of 36 post-mortem lenses of humans aged 20 to 30 years, showed relatively even transmission from 440 to 600 nm but a strong reduction in transmittance below 440 nm (Brainard et al., 1997). Since mean  $\pm$  SEM percent lens transmittance at 420 and 460 nm was  $37.23 \pm 7.88$  and  $56.33 \pm 10.1$ , corneal light irradiances at 420 nm were adjusted to compensate for reduced stimulus transmission to the retina in both studies.

### Statistics

Two-tailed Student *t* tests were used to assess significance of raw melatonin change. The melatonin data were then converted to percent control-adjusted melatonin change scores (Brainard et al., 2001). Sets of preexposure melatonin values and percent control-adjusted melatonin change scores were analyzed with one-way, repeated-measures analysis of variance (ANOVA). Significant differences between groups were assessed with the post hoc Fisher protected least significant difference (PLSD) test with alpha at 0.05. A fluence-response curve was fit to a 4-parameter model for the mean percent control-adjusted melatonin

suppression data. The formula for this curve includes factors derived from earlier work on the melatonin suppression action spectrum (Brainard et al., 2001). Fit of the data to the curve was assessed by coefficient of correlation.

## RESULTS

The fluence-response data are illustrated in Figures 1 and 2. There were no significant differences ( $F = 0.69$ ,  $df = 8$ ,  $p = 0.70$ ) between sets of preexposure melatonin values, indicating that 2:00 AM plasma levels were consistent across all of the study nights. Figure 1 shows the mean  $\pm$  SEM pre- and postexposure melatonin values. Paired Students  $t$  tests showed significant melatonin suppression by retinal irradiances at or above 11  $\mu\text{W}/\text{cm}^2$ . All melatonin data were converted to control-adjusted percent change scores (Brainard et al., 2001), and ANOVA showed a significant effect of retinal light intensity on melatonin percent control-adjusted change scores ( $F = 11.74$ ,  $p < 0.0001$ ). Post hoc Fisher PLSD tests demonstrated that compared with the lowest irradiance of 0.016  $\mu\text{W}/\text{cm}^2$ , intensities at or above 4.1  $\mu\text{W}/\text{cm}^2$  significantly suppressed melatonin. In all cases, irradiances above 4.1  $\mu\text{W}/\text{cm}^2$  were significantly stronger in suppressing melatonin compared with the irradiances 2 steps lower. Figure 2 illustrates a sigmoidal fluence-response curve plotting melatonin percent control-adjusted scores against photon density. The curve formula is inset in the figure ( $R^2 = 0.93$ ).

In the wavelength comparison study, for the 420-nm and 460-nm light exposure and the dark control nights, mean preexposure raw melatonin values were  $73.9 \pm 8.7$ ,  $68.3 \pm 7.6$ , and  $69.8 \pm 8.6$  pg/mL, respectively. There were no significant differences ( $F = 1.27$ ,  $df = 2$ ,  $p = 0.29$ ) across these values, indicating that melatonin levels were consistent across all study nights. Mean postexposure scores were  $84.4 \pm 10.4$ ,  $76.1 \pm 9.5$ , and  $56.6 \pm 8.8$  pg/mL, respectively. Melatonin did not change significantly relative to the 420-nm exposure ( $t = 0.34$ ,  $df = 17$ ,  $p = 0.74$ ), decreased significantly with the 460-nm exposure ( $t = 2.25$ ,  $df = 17$ ,  $p < 0.04$ ), and increased significantly during the control night ( $t = -3.32$ ,  $df = 17$ ,  $p < 0.001$ ). For direct comparison of responses to 420 and 460 nm, Figure 3 illustrates percent control-adjusted melatonin suppression at equal retinal photon densities. These data reveal that 460 nm is significantly stronger than 420 nm in suppressing melatonin ( $t = 2.3$ ,  $df = 17$ ,  $p < 0.04$ ), although 5 of the 19 subjects had a greater melatonin suppression response to 420-nm versus 460-nm light. In this study,

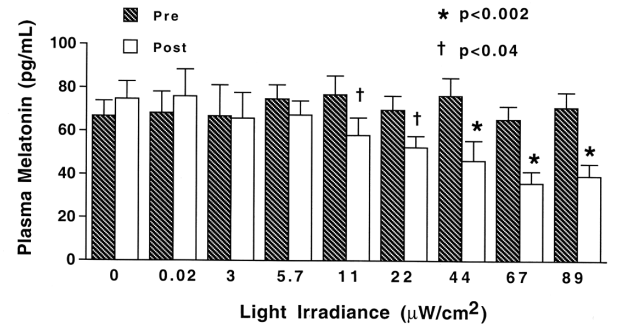


Figure 1. In this graph, bars represent group mean  $\pm$  SEM melatonin values ( $n = 8$ ) before and after monochromatic light exposure at 420 nm. Analysis of variance and post hoc Fisher protected least significant difference tests demonstrated which retinal light intensities significantly suppressed melatonin.

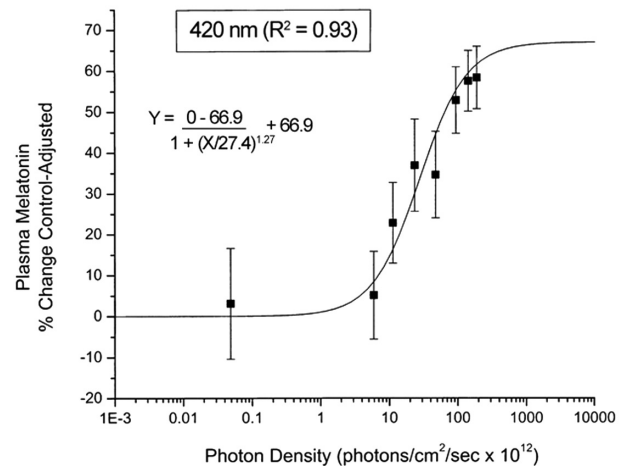


Figure 2. This figure demonstrates the fitted fluence-response curve for retinal irradiance photon density and percent control-adjusted melatonin suppression on a semilog scale ( $n = 8$ ). Each data point represents 1 group mean  $\pm$  SEM. The curve is consistent with the 8 fluence-response curves for melatonin suppression with monochromatic light between 440 and 600 nm (Brainard et al., 2001).

there was good repeatability in mean  $\pm$  SEM melatonin suppression responses compared with the fluence-response experiments. In the published 460-nm fluence-response curve (Brainard et al., 2001),  $1.21 \times 10^{13}$  photons/ $\text{cm}^2/\text{sec}$  elicited a  $45.3\% \pm 11.5\%$  control-adjusted melatonin suppression, while in this study, that photon dose elicited a  $44.4\% \pm 9.1\%$  control-adjusted melatonin suppression. Similarly, in the 420-nm fluence-response curve described above and in this study,  $1.21 \times 10^{13}$  photons/ $\text{cm}^2/\text{sec}$  exposure elicited a  $22.8\% \pm 9.7\%$  and  $20.2\% \pm 9.1\%$  control-adjusted melatonin suppression, respectively.

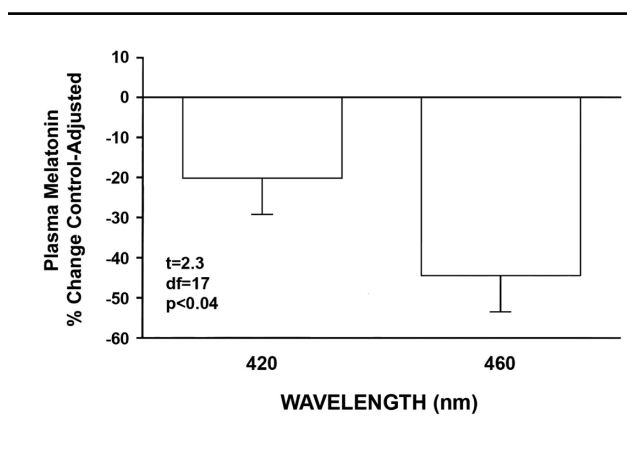


Figure 3. The bars represent group mean  $\pm$  SEM values relative to an equal photon dose of  $1.21 \times 10^{13}$  photons/cm<sup>2</sup> of retinal irradiance. These data show that the 460-nm percent control-adjusted plasma melatonin suppression is significantly stronger than that for 420 nm.

## DISCUSSION

The present data demonstrate a full fluence-response relationship between 420-nm exposure and melatonin suppression in humans. The 420-nm curve is consistent with the fluence-response curves for 8 other monochromatic wavelengths in the original melatonin suppression action spectrum (Brainard et al., 2001). In that action spectrum, a 420-nm half-saturation constant of  $18.3 \times 10^{12}$  photons/cm<sup>2</sup>/sec was estimated from a very limited data set. That estimate was reasonably consistent but lower than the half-saturation constant of  $27.4 \times 10^{12}$  photons/cm<sup>2</sup>/sec derived from the complete fluence-response curve. This new 420-nm data point has a better fit to the original action spectrum model but does not change the calculated peak (464 nm, 446-477 nm  $\pm$  1 SD) since the curve-fitting method is based on the long-wavelength limb of sensitivity of the action spectrum (Partridge and DeGrip, 1991). The within-subjects comparison showed that 460-nm light is significantly stronger than 420-nm light for suppressing melatonin. These results clarify the short visible wavelength sensitivity of the human melatonin suppression action spectrum.

The acute light-induced melatonin suppression response is a broadly used indicator for photic input to the SCN and has been used to elucidate the ocular and neural physiology for circadian and neuroendocrine regulation (CIE, 2004, 2006; Brainard et al., 1997). Although full-analytic action spectra have yet to be developed, a set of studies has confirmed that blue monochromatic light is more potent than other wavelengths for evoking circadian phase shifts and

enhancing acute alertness in humans (Lockley et al., 2003; Cajochen et al., 2005). Together, those results are consistent with the 9 more fully developed analytic action spectra for circadian and neuroendocrine responses (for review, see Brainard and Hanifin, 2005).

The 420-nm results are consistent with the results of 2 in vitro studies in which cells transfected with the melanopsin gene exhibit peak photosensitivities at 479 nm and 480 nm (Qiu et al., 2005; Panda et al., 2005). In addition, studies on amphioxus melanopsin show a peak absorbance near 485 nm (Koyanagi et al., 2005). Together, the 9 in vivo action spectra and the 3 in vitro studies indicate peak sensitivity in the blue part of the spectrum. In contrast, 2 in vitro studies show peaks in the violet-indigo-ultraviolet parts of the spectrum. Specifically, mouse cells transfected with the melanopsin gene have a peak photosensitivity in the range of 360 to 430 nm, and the direct absorption spectrum for melanopsin showed the strongest activation by 420- to 440-nm light (Melyan et al., 2005; Newman et al., 2003). This discrepancy may be due to the difference of in vitro melanopsin responsiveness by itself or in a given cell type versus its in vivo presence in ipRGCs that are closely connected to other retinal cells. There is increasing evidence that melanopsin may function as an invertebrate-like, bistable photopigment with both sensory and regenerative functions that have differing peaks of wavelength sensitivity (Koyanagi et al., 2005; Mure et al., 2007; Rollag, 2008 [this issue]). Hence, some in vitro systems may not match the systemic action spectra due to the blending of different melanopsin isomerization states. Further work is needed to clarify how the ipRGC-melanopsin system supports the wavelength sensitivity of systemic action spectra.

Despite abundant evidence that the melanopsin-containing ipRGCs provide primary input for circadian and neuroendocrine phototransduction, the rod and cone photoreceptors still play a role in this physiology. Melanopsin- and cone-knockout mice show that the classical visual photoreceptors can compensate for the loss of melanopsin and at least partially mediate light-induced circadian, neuroendocrine, and neurobehavioral responses (Panda et al., 2002; Lucas et al., 2003; Dkhissi-Benyahya et al., 2007). In contrast, when both melanopsin is knocked out and the rods and cones are compromised, animals lose all visual and nonvisual photoreceptive functions of the eye (Hattar et al., 2003; Panda et al., 2003). Furthermore, cellular recording studies in nonhuman primate retinas have demonstrated that

rod and cone cells can directly activate ipRGCs (Dacey et al., 2005). Data from human studies suggest that the visual rods and cones may provide input to the SCN (Hebert et al., 2002; Figueiro et al., 2004; Jasser et al., 2006; Revell and Skene, 2007). It is important to recognize that despite rapid experimental progress on ipRGC physiology, it is currently unknown how these newly discovered photoreceptors work with the classical visual photoreceptors in transducing light in the dynamic, complex polychromatic environments where humans carry out their daily activities.

Importantly, in working with short wavelengths such as 420 nm, there is the potential for significant radiometric measurement error (American National Standards Institute and Illuminating Engineering Society of North America 2001). Special care is required in calibrating and benchmarking meters for accurately quantifying short-wavelength visible light. Furthermore, it is critical that the measured light stimuli represent the stimuli reaching the relevant photopigments. Human factors that can modify the measured stimulus include head and eye motion, squinting and eye closure, pupillary reflexes, and ocular media light transduction (Brainard et al., 1997). Most of these factors are controlled in the exposure techniques reported here. In ocular media light transmission, the cornea and aqueous and vitreous humors normally transmit nearly 100% of visible wavelengths to the retina. In contrast, as the human lens ages, it develops pigmentation that attenuates shorter visible wavelength transmission (Brainard et al., 1997; Pokorny et al., 1987). In this study, restricting the age of volunteers to 18 to 30 years partially controlled this factor. Measurements of transmittance of 36 postmortem human lenses in this age range showed relatively even transmission from 440 to 600 nm. Compared with lens transmission at 460 nm, however, there was a mean 45% reduction in transmission at 420 nm (Brainard et al., 1997). Thus, measured corneal light irradiances at 420 nm were adjusted to compensate for this reduced transmission. Such adjustments are advisable for all studies using short-wavelength visible light. One study that used 456-nm light showed reduced melatonin suppression in older versus younger women (mean ages  $57 \pm 5$  and  $24 \pm 3$  years), suggesting that the sensitivity loss was likely due to age-related changes in subjects' lenses (Herljevic et al., 2005).

Photobiological hazards such as infrared and ultraviolet cataract, photokeratitis, photoretinitis, and ultraviolet erythema have been identified relative to overexposure of the skin and eyes to the ultraviolet,

visible, and infrared spectra. Whether using short-wavelength light experimentally or for pragmatic purposes, it is important to verify that exposures fall within established national and international safety limits (CIE, 2002; American Conference of Governmental Industrial Hygienists [ACGIH], 2006). The short-wavelength stimuli used in this study were well within the established ocular safety limits. Although the ACGIH standards are updated yearly based on the current published literature, some investigators debate if these standards are sufficiently stringent.

A wealth of data published in the past 25 years has demonstrated that light can be a potent biological, behavioral, and therapeutic stimulus in humans (CIE, 2004, 2006). The data presented here extend our understanding of the wavelength sensitivity of the photoreceptor system that serves as the input system for nonvisual, neurobehavioral regulation in humans. Industrialized societies employ light extensively in both public and private buildings to support vision, visual comfort, and aesthetic appreciation within these environments. Since light is also a potent regulator of human circadian and neuroendocrine physiology and different photoreceptive systems mediate visual and neurobehavioral responses, future lighting strategies will need to provide illumination for human neurobehavioral regulation as well as vision. Collectively, lighting manufacturers, lighting designers, and architectural engineers have opened the door to understanding this physiology and are considering the development of applications stemming from these discoveries (CIE, 2004, 2006). Indeed, the aerospace community is exploring how lighting can be used to support vision, circadian regulation, and alertness of astronauts in advanced human environments such as the International Space Station and the planned lunar habitat (Dijk et al., 2001; Gronfier et al., 2007).

## ACKNOWLEDGMENTS

The authors are grateful for the expert assistance of S. P. Wengraitis, Laser/Optical Radiation Program, Aberdeen Proving Ground, Maryland, for radiometric calibration; the technical and logistical assistance from C. Alocillo, P. Becerra, J. Cooke, W. Coyle, R. L. Fucci Jr., J. Gardner, R. Glasgow, R. Kovach, and J. Monnier; and editorial, referencing, administrative, and graphic assistance from D. L. Brainard, M. Jablonski, C. Penrose, and B. Warfield. Preliminary reports on this work were given at the following meetings: Soc Light Treatment Biol Rhythms 2001,

Gordon Res Conf 2002, Soc Res Biol Rhythms 2002 and 2004, Bioastronautics 2003 and 2005, and Humans in Space 2005. Grant support was from the National Space Biomedical Research Institute through NASA NCC 9-58, NIH RO1NS36590, NSF IBN9809916, and the Phil Sect IESNA. Inspiration for this work came from the 281 series of the Edgar Cayce readings.

## REFERENCES

- American Conference of Governmental Industrial Hygienists (2006) Nonionizing radiation and fields. In *Documentation of the Threshold Limit Values and Biological Exposure Indices*, Cincinnati, OH, American Conference of Governmental Industrial Hygienists.
- American National Standards Institute and Illuminating Engineering Society of North America (2001) *Recommended Practice for Photobiological Safety for Lamps and Lamp Systems- Measurement Techniques RP-27.2*. New York: Illuminating Engineering Society of North America.
- Berson DM, Dunn FA, and Takao M (2002) Phototransduction by retinal ganglion cells that set the circadian clock. *Science* 295:1070-1073.
- Brainard GC and Hanifin JP (2005) Photons, clocks and consciousness. *J Biol Rhythms* 20:314-325.
- Brainard GC, Hanifin JP, Greeson JM, Byrne B, Glickman G, Gerner E, and Rollag MD (2001) Action spectrum for melatonin regulation in humans: Evidence for a novel circadian photoreceptor. *J Neurosci* 21:6405-6412.
- Brainard GC, Rollag MD, and Hanifin JP (1997) Photic regulation of melatonin in humans: Ocular and neural signal transduction. *J Biol Rhythms* 12:537-546.
- Cajochen C, Munch M, Koblalka S, Krauchi K, Steiner R, Oelhafen P, Orgul S, and Wirz-Justice A (2005) High sensitivity of human melatonin, alertness, thermoregulation, and heart rate to short wavelength light. *J Clin Endocrinol Metab* 90:1311-1316.
- Commission Internationale de l'Eclairage (2002) *Photobiological Safety of Lamps and Lamp Systems*. CIE Pub. No. S 009/E:2002. Vienna: Commission Internationale de l'Eclairage.
- Commission Internationale de l'Eclairage (2004) *Ocular Lighting Effects on Human Physiology and Behaviour*. Vienna: Commission Internationale de l'Eclairage.
- Commission Internationale de l'Eclairage (2006) *Proceedings of the 2nd CIE Symposium on Light and Health*. Vienna: Commission Internationale de l'Eclairage.
- Dacey DM, Liao H-W, Peterson BB, Robinson FR, Smith VC, Pokorny J, Yau K-W, and Gamlin PD (2005) Melanopsin-expressing ganglion cells in primate retina signal colour and irradiance and project to the LGN. *Nature* 433:749-754.
- Dijk DJ, Neri DF, Wyatt JK, Ronda JM, Riel E, Ritz-De Cecco A, Hughes RJ, Elliott AR, Prisk GK, West JB, et al. (2001) Sleep, performance, circadian rhythms, and light-dark cycles during two space shuttle flights. *Am J Physiol* 281:R1647-R1664.
- Dkhissi-Benyahya O, Gronfier C, De Vanssay W, Flamant F, and Cooper HM (2007) Modeling the role of mid-wavelength cones in circadian responses to light. *Neuron* 53:677-687.
- Figueiro MG, Bullough JD, Parsons RH, and Rea MS (2004) Preliminary evidence for spectral opponency in the suppression of melatonin by light in humans. *Neuroreport* 15:313-316.
- Gamlin PDR, McDougal DH, Pokorny J, Smith VC, Yau K-W, and Dacey DM (2007) Human and macaque pupil responses driven by melanopsin-containing retinal ganglion cells. *Vision Res* 47:946-954.
- Gooley JJ, Lu J, Chou TC, Scammell TE, and Saper CB (2001) Melanopsin in cell of origin of the retinohypothalamic tract. *Nature Neurosci* 4:1165.
- Gronfier C, Wright KP, Kronauer RE, and Czeisler CA (2007) Entrainment of the human circadian pacemaker to longer-than-24-h days. *Proc Natl Acad Sci USA* 104:9081-9086.
- Hattar S, Liao H-W, Takao M, Berson DM, and Yau K-W (2002) Melanopsin-containing retinal ganglion cells: Architecture, projections, and intrinsic photosensitivity. *Science* 295:1065-1070.
- Hattar S, Lucas RJ, Mrosovsky N, Thompson S, Douglas RH, Hankins MW, Lem J, Biel M, Hofmann F, Foster RG, et al. (2003) Melanopsin and rod-cone photoreceptive systems account for all major accessory visual functions in mice. *Nature* 424:76-81.
- Hebert M, Martin SK, Lee C, and Eastman CI (2002) The effects of prior light history on the suppression of melatonin by light in humans. *J Pineal Res* 33:198-203.
- Herljevic M, Middleton B, Thapan K, and Skene DJ (2005) Light-induced melatonin suppression: Age-related reduction in response to short wavelength light. *Exp Gerontol* 40:237-242.
- Jasser SA, Hanifin JP, Rollag MD, and Brainard GC (2006) Dim light adaptation attenuates acute melatonin suppression in humans. *J Biol Rhythms* 21:394-404.
- Koyanagi M, Kubokawa K, Tsukamoto H, Shichida Y, and Terakita A (2005) Cephalochordate melanopsin: Evolutionary linkage between invertebrate visual cells and vertebrate photosensitive retinal ganglion cells. *Curr Biol* 15:1065-1069.
- Lockley SW, Brainard GC, and Czeisler CA (2003) High sensitivity of the human circadian melatonin rhythm to resetting by short wavelength light. *J Clin Endocr Metab* 88:4502-4505.
- Lucas RJ, Hattar S, Takao M, Berson DM, Foster RG, and Yau KW (2003) Diminished pupillary light reflex at high irradiances in melanopsin-knockout mice. *Science* 299:245-247.
- Melyan Z, Tarttelin EE, Bellingham J, Lucas RJ, and Hankins MW (2005) Addition of human melanopsin renders mammalian cells photoresponsive. *Nature* 433:741-745.
- Mure LS, Rieux C, Hattar S, and Cooper HM (2007) Melanopsin-dependent nonvisual responses: Evidence for photopigment bistability in vivo. *J Biol Rhythms* 22:411-424.
- Newman LA, Walker MT, Brown RL, Cronin TW, and Robinson PR (2003) Melanopsin forms a functional short-wavelength photopigment. *Biochemistry* 42:12734-12738.
- Panda S, Nayak SK, Campo B, Walker JR, Hogenesch JB, and Jegla T (2005) Illumination of melatonin signaling pathway. *Science* 307:600-604.
- Panda S, Provencio I, Tu DC, Pires SS, Rollag MD, Castrucci AM, Pletcher MT, Sato TK, Wiltshire T, Andahazy M,

- et al. (2003) Melanopsin is required for non-image-forming photic responses in blind mice. *Science* 301:525-527.
- Panda S, Sato TK, Castrucci AM, Rollag MD, DeGrip WJ, Hogenesch JB, Provencio I, and Kay SA (2002) Melanopsin (Opn4) requirement for normal light-induced circadian phase-shifting. *Science* 298:2213-2216.
- Partridge JC and De Grip WJ (1991) A new template for rhodopsin (vitamin A1 based) visual pigments. *Vision Res* 31:619-630.
- Pokorny J, Smith VC, and Lutze M (1987) Aging of the human lens. *Appl Optics* 26:1437-1440.
- Provencio I, Rodriguez IR, Jiang G, Hayes WP, Moreira EF, and Rollag MD (2000) A novel human opsin in the inner retina. *J Neurosci* 20:600-605.
- Qiu X, Kumbalasisri T, Carlson SM, Wong KY, Krishna V, Provencio I, and Berson D (2005) Induction of photosensitivity by heterologous expression of melatonin. *Nature* 433:745-749.
- Revell VL and Skene DJ (2007) Light-induced melatonin suppression in humans with polychromatic and monochromatic light. *Chronobiol Int* 24:1125-1137.
- Rollag MD (2008) Does melanopsin bistability have physiological consequences? *J Biol Rhythms* **IN PRESS**.
- Thapan K, Arendt J, and Skene DJ (2001) An action spectrum for melatonin suppression: Evidence for a novel non-rod, non-cone photoreceptor system in humans. *J Physiol* 535:261-267.